

Metallopeptoids†

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***N*-substituted glycine peptoid oligomers bearing hydroxyquinoline ligands form complexes with Cu(II) and Co(II) in which the chiral helical secondary structure of the foldamers is enhanced upon metal binding and establishes a stereogenic environment for metal coordination.**

Peptoids—oligomers of *N*-substituted glycine—have emerged as intriguing mimics of polypeptides, particularly with respect to their ability to form well-defined folded architectures. Although peptoids are incapable of forming hydrogen-bonded networks along the backbone, many peptoid sequences exhibit a remarkable propensity for folding at small oligomer chain lengths, a property that has been ascribed to the influence of local steric and stereoelectronic interactions.¹ Peptoid oligomers can be synthesized efficiently by solid-phase methods, allowing the facile introduction of a variety of side chains within the context of well-defined asymmetric environments. This feature enables the coordinated display of multiple chemical functionalities, potentially emulating binding sites and enzymatic sites in proteins. The conformation and attendant catalytic activity of proteins and polypeptides, however, often depends upon the coordination of metal species.² Recent studies have investigated the interaction of metal ions with biomimetic oligomers (a.k.a. foldamers)^{3,4} and include peptoids that bind zinc.⁵ Further advances in mimicking natural systems require an improved understanding of the reciprocating influences of metal coordination and peptoid secondary structure. We have demonstrated that multidentate metal-binding ligands can be introduced as pendant groups in peptoid sequences.⁶ Herein we report the formation of metal complexes using these ligands, wherein helical secondary structure is enhanced upon metal binding and enforces a chiral environment about the metal center.

In order to explore the influence of metal binding on the conformation of chiral peptoids, hydroxyquinoline ligands were introduced as *N*-substituted pendant groups on glycine oligomer scaffolds. Chiral (*S*)-1-phenylethyl groups (*N*spe) were introduced at other *N*-positions to enforce helicity resembling that of the polypro-type I helix, with a pitch of roughly three residues per turn.^{1a} The hydroxyquinoline ligands were expected to bind divalent metal ions, producing tetra-coordinated metal species.⁷ Specifically, **H₁₅** and **H₂₆** (Fig. 1; H = hydroxyquinoline; 5 = pentamer, 6 = hexamer)

were synthesized as model systems for comparison of inter- and intramolecular metal complex formation, respectively. These compounds were chosen because each contains four bulky chiral side chains, which are sufficient for achieving helicity in oligomers of this length.^{1c} The pentamer **H₁₅**, with one hydroxyquinoline site at the *N*-terminus, was expected to form a peptoid duplex upon metal binding (2 : 1 peptoid:metal). The hexamer **H₂₆** contains two hydroxyquinoline ligands, endowing this oligomer with the capacity to form an intramolecular 1 : 1 peptoid:metal complex (Fig. 1). The disposition of the ligands at positions *i* and *i* + 3 in the sequence matches the pitch of the helix and is designed to orient these groups in proximity on the same face of the scaffold, separated by one helical turn.

Oligomers **H₁₅** and **H₂₆** were synthesized using solid phase methods, with 8-hydroxy-2-quinolinemethylamine and (*S*)-(-)-1-phenylethylamine as synthons, employing “submonomer” protocols.^{6,8} The peptoids were cleaved from the solid support and purified by HPLC, with no detectable impurities (>98% purity). The molecular weights measured by electrospray mass spectrometry were consistent with the masses expected for **H₁₅** and **H₂₆**.

Metal-free **H₁₅** and **H₂₆** exhibited absorption bands near $\lambda = 245$ nm and 307 nm in 4 : 1 methanol:water (the solvent of choice for solubility and spectral transparency). Upon addition of cupric acetate, binding of Cu²⁺ produced new absorption bands at $\lambda = 260, 366$ nm and $\lambda = 263, 384$ nm for **H₁₅** and **H₂₆**, respectively (Fig. 2). Job plots constructed from UV titration of the peptoids with cupric acetate were consistent with 2 : 1 (**H₁₅**)₂Cu duplex and a 1 : 1 (**H₂₆**)Cu complex that signaled the formation of an intramolecular tetra-coordinated complex. Likewise, new absorption bands at $\lambda = 262, 365$ nm

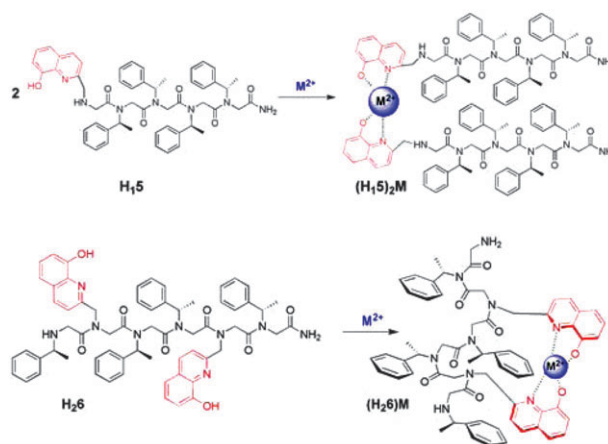


Fig. 1 The expected 2:1 and 1:1 peptoid-metal complexes formed from **H₁₅** and **H₂₆**, respectively.

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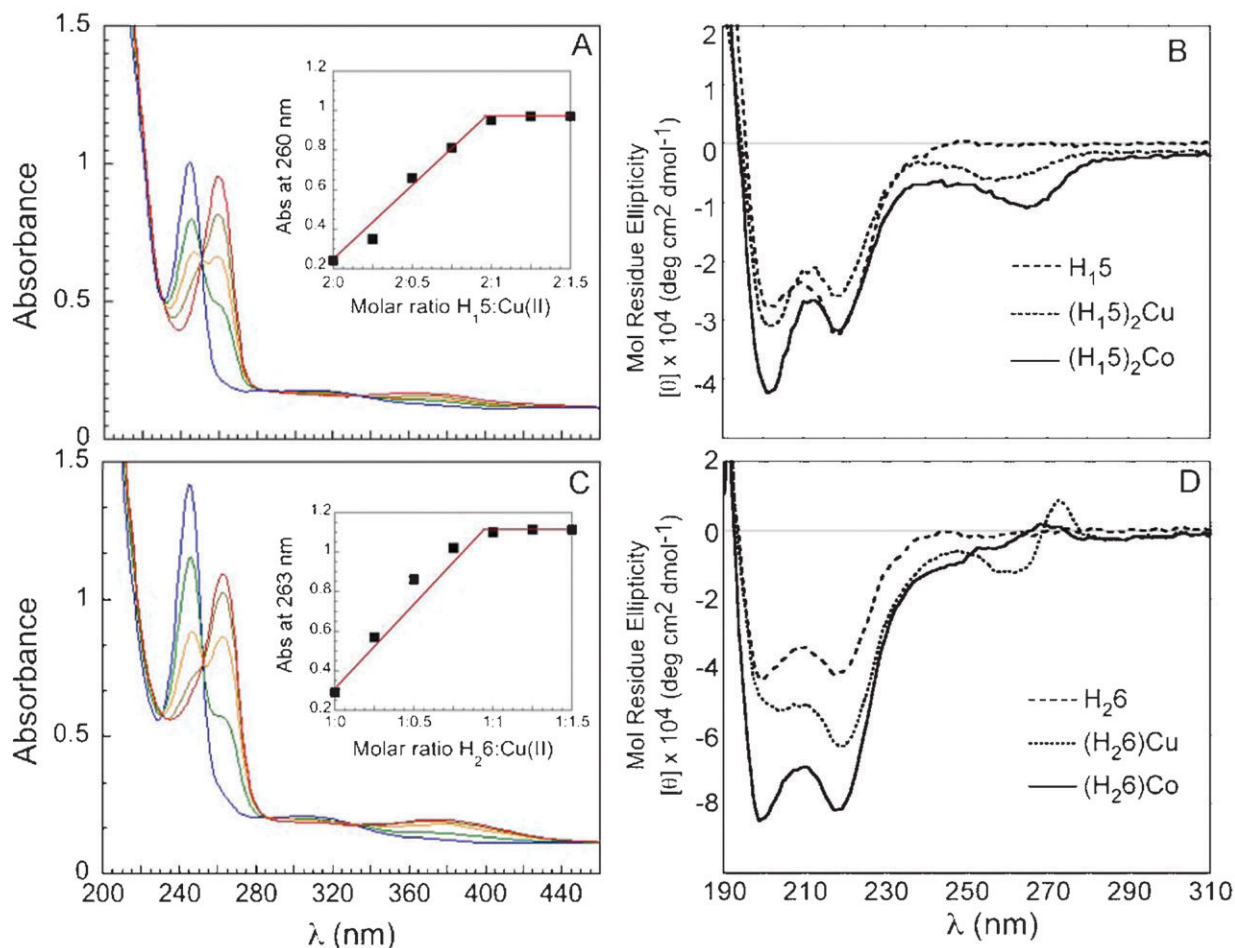


Fig. 2 (A) UV-Vis spectra and Job plot for titration of **H15** with Cu^{2+} . (B) CD spectra for (**H15**), (**H15**)₂Cu, and (**H15**)₂Co. (C) UV-Vis spectra and Job plot for titration of **H26** with Cu^{2+} . (D) CD spectra for (**H26**), (**H26**)Cu, and (**H26**)Co. UV-Vis spectra: (A) 17 μM peptoid in MeOH : H₂O (4 : 1) solution, (C) 40 μM peptoid in MeOH : H₂O (4 : 1) solution. The peptoid solutions were titrated with 2 μL aliquots of a metal ion (5 mM in H₂O), in multiple steps (blue = free ligand, red = metal complex); CD spectra: 100 μM MeOH : H₂O (4 : 1) solutions.

and $\lambda = 266, 375 \text{ nm}$ appeared upon addition of cobalt(II) acetate. The corresponding Job plots from UV titration revealed the formation of 2 : 1 (**H15**)₂Co and 1 : 1 (**H26**)Co. The peptoid-to-metal ratio was corroborated further by electro-spray mass spectrometry and MALDI-TOF (Table S1†). No evidence for the formation of higher order complexes with **H26** (e.g. 2 : 2 complexes) was detected. The association constants for the two-step equilibrium responsible for formation of metal complexes with both peptoids were estimated from the UV titration experiments.⁹ The values for (**H15**)₂Cu, as calculated by nonlinear regression curve fitting² (see ESI†) are $\beta_1 (K_1) = 3.75 \pm 0.23 \times 10^5 \text{ M}^{-1}$ and $\beta_2 (K_1 \cdot K_2) = 5.45 \pm 0.36 \times 10^9 \text{ M}^{-2}$. The corresponding values for (**H15**)₂Co are $\beta_1 = 7.4 \pm 1.7 \times 10^5 \text{ M}^{-1}$ and $\beta_2 = 9.45 \pm 2.2 \times 10^9 \text{ M}^{-2}$. The association constants for (**H26**)Cu and (**H26**)Co reflected much stronger binding, with an estimated value of $K_1 > 10^{14}$ and $K_1 = 1.36 \pm 0.18 \times 10^{13} \text{ M}^{-1}$, respectively. The larger binding constant for the (**H26**)M complexes suggests the existence of a preorganized binding environment due to the disposition of the hydroxyquinoline ligands on the helical backbone (see below). Indeed, differences in affinity that arise from a preorganization of the oligomer sequence have been shown to yield higher association

constants in Zn^{2+} -binding peptides.¹⁰ Moreover, the association constants for an unstructured achiral analog of (**H26**)Co incorporating *N*-methoxyethyl monomers (see ESI†), reflected weaker binding in comparison with the chiral **H26** complex, with a value of $K_1 = 6.4 \pm 2.2 \times 10^{11} \text{ M}^{-1}$.

Circular dichroism (CD) measurements revealed substantial changes upon complex formation. Solutions of the metal complexes exhibited increases, relative to the metal-free peptoids, in the magnitude of the CD signals near 200 nm and 220 nm. In the case of (**H15**)₂M this suggests an increase in conformational order, as the magnitude of the signal reflects the degree of helicity. Similar changes in CD spectra have been reported for peptide bundles formed upon metal complexation.^{2,11} Notably, no changes in CD intensity were observed for metal complexes of peptoid bundles with monodentate ligands.⁵ The increase in the CD signal was more dramatic for (**H26**)M, consistent with greater conformational constraint and enhanced secondary structure content due to intramolecular metal complexation.^{12,13} Metal binding to **H15** and **H26** also produced new CD peaks between 240 and 280 nm, the region corresponding to the 8-hydroxyquinoline π - π^* transition, which reflects the transmission of the stereogenic character of the peptoid scaffold to the metal

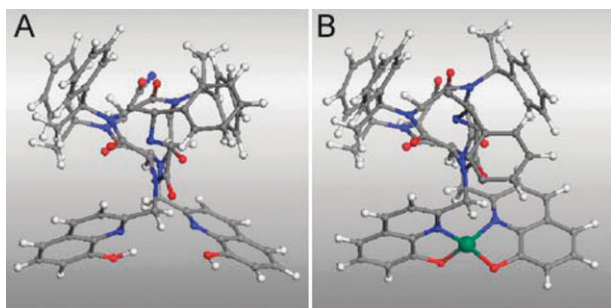


Fig. 3 Energy-minimized structures of (A) metal-free **H₂₆** and (B) (**H₂₆**)Cu.^{20,21} Hydrogen atoms within the helix are omitted for clarity.

center.^{2,14,15} These results indicate the reciprocating effects of metal binding—the chirality of the peptoid backbone establishes an asymmetric environment about the metal center while metal complexation enhances the helical character of the backbone.

Notably, Cu(hydroxyquinolate)₂ adopts a trans-planar configuration (*i.e.* square planar with O trans to O and N trans to N).¹⁶ The trans-planar configuration is not possible in the intramolecular (**H₂₆**)M complexes because of the parallel projection of the two hydroxyquinoline ligands from the peptoid scaffold and steric interactions that prohibit a square planar geometry. Consequently, the peptoid scaffold in **H₂₆** enforces an unusual coordination environment while transmitting its stereogenic character to the metal site. The metal center most likely adopts local C₂ symmetry (*i.e.* pseudotetrahedral) about the Cu(II) and Co(II) centers. This is supported by molecular modeling, which revealed that formation of a pseudotetrahedral metal complex in (**H₂₆**)Cu can be achieved without substantial repositioning of the ligands compared with the metal-free structure (Fig. 3). Moreover, the existence of this preorganized binding environment is consistent with the very large binding constants observed for the 1 : 1 complexes.

In contrast, a trans-planar configuration should not be sterically prohibited in the unconstrained intermolecular (**H₁₅**)₂M complexes, which also exhibited CD signals in the π - π^* region. The 8-hydroxyquinoline π - π^* CD signal for (**H₂₆**)Cu exhibited an exciton couplet,^{17–19} with a minimum at 260 nm and a maximum at 273 nm, crossing $\epsilon = 0$ near 268 nm. In contrast, (**H₂₆**)Co exhibited a very weak CD signal in this region with a corresponding weak excitonic couplet. The absence of a similar couplet for the intermolecular (**H₁₅**)₂Cu and (**H₁₅**)₂Co complexes and the close agreement between the CD minima and their respective λ_{max} values, however, indicates the absence of exciton coupling. This most likely reflects reduced conformational constraints in the intermolecular complexes, which can diminish the effect of the dipole-dipole interactions responsible for an exciton couplet.

Collectively, these observations reveal that metal binding in designed peptoid sequences can introduce conformational constraints and chiral environments about the metal centers. We anticipate that this strategy can be extended to the synthesis of peptoid podands and other foldamers with unique secondary, tertiary or quaternary structures, promising

new architectures with utility for asymmetric catalysis and materials science.

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